

RENAL NERVE STIMULATION AS A POTENTIAL ELECTROCEUTICAL THERAPY FOR OBSTRUCTIVE SLEEP APNEA

Ibrahim M. Salman ^{1,2}, Omar Z. Ameer ¹, Ghaith K. Mansour ^{1*}, Hussam N. Alhamad ¹ and Stephen J. Lewis ²

¹ Department of Pharmaceutical Sciences, College of Pharmacy, Alfaisal University, Riyadh, Saudi Arabia;

² Department of Pediatrics, School of Medicine, Case Western Reserve University, Cleveland, Ohio, USA

Introduction

- Obstructive sleep apnea (OSA) is a prevalent sleep-related breathing disorder (Fig. 1) marked by recurrent upper airway collapse during sleep, resulting in sleep disruption, reduced oxygen saturation and an increased risk of mortality and cardiovascular events [1].
- The role of the renal nerves (Fig. 2) in blood pressure control is well-established [2]. However, their role in modulating respiration remains largely unexplored.
- Our recent data in spontaneously hypertensive rats (SHRs) suggest that lowintensity renal nerve stimulation (RNS) can enhance respiratory functions [3].
- Neuromodulation of the renal nerves may therefore offer a therapeutic alternative for OSA or other breathing abnormalities.



obstructive sleep apnea.



 Hemodynamically, RNS evoked an immediate but reversible 33 ± 4 mmHg $(27 \pm 3\%)$ reduction in MAP (**Fig. 7**).



Fig. 7: Effect of renal nerve stimulation (RNS) on mean arterial pressure (MAP) in Zucker fat rats (ZFRs). Data were mean ± SEM. ***P<0.001 vs. baseline.

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• RNS increased RR by 7 ± 2 breaths.min⁻¹ (7 ± 2%), with effects persisting temporarily beyond stimulation (Fig. 8).



Fig. 8: Effect of renal nerve stimulation (RNS) on respiration rate (RR) in Zucker fat rats (ZFRs). Data were mean ± SEM. *P<0.05, ***P<0.001 vs. baseline.

• RNS did not alter overall AP. However, UAP dropped by 0.6 ± 0.2 mmHg (3 ± 1%) during stimulation and remained relatively low after stimulation (Fig. 9).





Fig. 9: Effect of renal nerve stimulation (RNS) on upper airway pressure (UAP) in Zucker fat rats

Results

 This study aimed to investigate ventilatory responses to RNS in a genetic model of obesitydriven OSA (Fig. 3), the Zucker fat rat (ZFR).

Fig. 3: The Zucker fat rat (ZFR) model of obesity and obstructive sleep apnea.

(ZFRs). Data were mean ± SEM. *P<0.05, ***P<0.001 vs. baseline. Baseline Stim Poststim Time (s) • RNS increased tidal AF by 0.04 \pm 0.01 ml.min⁻¹ (16 \pm 3%) during stimulation, with increases not instantly returning to baseline after stimulation (Fig. 10). 0.40 (_-ul: 0.35-0.30-(ml.min⁻¹ 0.3 Fig. 10: Effect of renal nerve stimulation (RNS) on tidal airway 0.2 flow (AF) in Zucker fat rats (ZFRs). AF ₩ ▼ 0.25 0.1 Data were mean ± SEM. **P<0.01, ***P<0.001 vs. baseline. *n* = 12 0.20 Baseline Stim Poststim 100 50 Time (s) • RNS lowered AR by 0.6 \pm 0.1 mmHg.min.ml⁻¹ (12 \pm 2%) when the stimulus was on, with the peak reduction briefly maintained when the stimulation was

- **Methods**
- **Animals:** Male ZFRs (*n* = 12, weight = 914 ± 24 g).
- Anaesthesia: Sodium pentobarbital (induction dose: 50 mg.kg⁻¹ i.p. and maintenance dose: 10 mg.kg.h⁻ ¹ i.v. delivered at 2 ml. h^{-1}).
- **RSN protocol:** Bipolar monophasic stimulation of the intact left renal nerve (5 Hz, 0.5 mA, 0.5 ms, 30s; Fig. 4–6) was repeated 5 times, averaged, and performed at 5-min intervals.
- Recorded output responses: Mean arterial pressure (MAP), upper airway pressure (UAP), airway pressure (AP), tidal air flow (AF), and diaphragmatic EMG (dEMG).
- Calculations: Respiration rate (RR) was derived from integrated dEMG signal, and airway resistance (AR) was







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Conclusions

off (Fig. 11).

- Our data suggest that the renal nerves are not only involved in the modulation of cardiovascular function, but their role also extends to central regulation of respiration.
- Neurostimulation of the renal nerves effectively enhances ventilatory responses in the ZFR model of obesity-associated OSA.

calculated by dividing AP values by corresponding AF measurements.

100 s

Fig. 6: Raw data Spike trace showing recorded physiological parameters.

Clinically, neuromodulation of the renal nerves may offer an alternative novel therapy for patients with OSA.

References

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* Contact

- Ghaith K. Mansour, Year 4 PharmD student
- Email: gkmansour@alfaisal.edu

